

Design of Peptides Using α,β -Dehydro-residues: Synthesis, Crystal Structure and Molecular Conformation of *N*-Boc-L-Val- Δ Phe- Δ Phe-L-Ala-OCH₃

SMITA BHATIA, SHARMISTHA DEY, PUNIT KAUR and TEJ P. SINGH

Department of Biophysics, All India Institute of Medical Sciences, New Delhi, India

Received 16 October 1995

Accepted 6 March 1996

Abstract: To obtain general rules of peptide design using α,β -dehydro-residues, a sequence with two consecutive Δ Phe-residues, Boc-L-Val- Δ Phe- Δ Phe-L-Ala-OCH₃, was synthesized by azlactone method in solution phase. The peptide was crystallized from its solution in an acetone/water mixture (70:30) in space group P6₁ with $a=b=14.912(3)$ Å, $c=25.548(5)$ Å, $V=4912.0(6)$ Å³. The structure was determined by direct methods and refined by a full matrix least-squares procedure to an *R* value of 0.079 for 2891 observed [$I \geq 3\sigma(I)$] reflections. The backbone torsion angles $\phi_1 = -54(1)^\circ$, $\psi_1 = 129(1)^\circ$, $\omega_1 = -177(1)^\circ$, $\phi_2 = 57(1)^\circ$, $\psi_2 = 15(1)^\circ$, $\omega_2 = -170(1)^\circ$, $\phi_3 = 80(1)^\circ$, $\psi_3 = 7(2)^\circ$, $\omega_3 = -177(1)^\circ$, $\phi_4 = -108(1)^\circ$ and $\psi_4^T = -34(1)^\circ$ suggest that the peptide adopts a folded conformation with two overlapping β -turns of types II and III'. These turns are stabilized by two intramolecular hydrogen bonds between the CO of the Boc group and the NH of Δ Phe³ and the CO of Val¹ and the NH of Ala⁴. The torsion angles of Δ Phe² and Δ Phe³ side chains are similar and indicate that the two Δ Phe residues are essentially planar. The folded molecules form head-to-tail intermolecular hydrogen bonds giving rise to continuous helical columns which run parallel to the *c*-axis. This structure established the formation of two β -turns of types II and III' respectively for sequences containing two consecutive Δ Phe residues at (*i*+2) and (*i*+3) positions with a branched β -carbon residue at one end of the tetrapeptide.

Keywords: β -turns; folded conformation; dehydro-residue; X-ray diffraction; consecutive dehydro-residue

INTRODUCTION

The α,β -unsaturated amino acid residues (dehydro- or Δ -residues) have been found to be strong inducers of folded conformations in peptides[1,2]. Previous studies have indicated that peptides containing two consecutive Δ Phe residues at the (*i*+1) and (*i*+2) positions of pseudotetrapeptides Ac- Δ Phe- Δ Phe-Gly[3] and Ac- Δ Phe- Δ Phe-Ala[4] generate an S-shaped structure with positive and negative (ϕ,ψ) torsion angles alternately. On the other hand it was shown that two consecutive Δ Phe residues at (*i*+2) and (*i*+3) positions in a pseudotetrapeptide Boc-

Ala- Δ Phe- Δ Phe-NHCH₃[5] adopt a conformation with two overlapping type III β -turns (incipient 3_{10} -helix). The recent investigations on Boc-Val- Δ Phe- Δ Phe-Val-OCH₃[6] with two consecutive Δ Phe residues at (*i*+2) and (*i*+3) positions indicated the formation of a folded structure with two overlapping β -turns of types II and III' respectively. The differences in the last two structures despite the identical positions of the two consecutive Δ Phe residues indicate that the Val residue is influenced differently when placed adjacent to a Δ Phe residue. This relates to the fact that the branched β -carbon residues such as Val and Ile have a strong conformational preference for side-chain orientation which is often staggered relative to the main chain. In view of the above differences in the folded conformations with two consecutive Δ Phe residues at the (*i*+2) and (*i*+3) positions, it is necessary to examine a sequence with a branched β -carbon residue on one side while any other residue is at the other end of two

Address for correspondence: Prof. T. P. Singh, Department of Biophysics, All India Institute of Medical Sciences, Ansari Nagar, New Delhi 110 029, India. Tel 0091-11-659-3201; Fax 0091-11-686-2263

consecutive Δ Phe residues. Therefore, we report here, the synthesis, crystal structure and molecular conformation of *N*-Boc-L-Val- Δ Phe- Δ Phe-L-Ala-OCH₃(1).

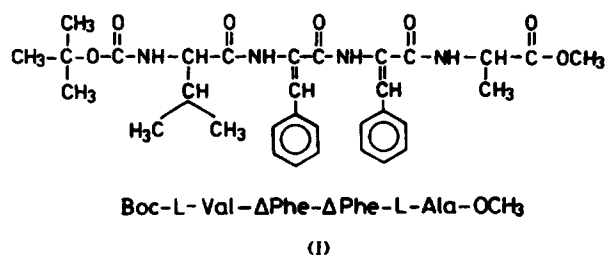
MATERIALS AND METHODS

Melting points recorded are uncorrected. Thin layer chromatography was carried out on silica gel G in solvent systems used (by volume): (a) CHCl₃:MeOH (9:1) and (b) *n*BuOH:AcOH:H₂O (4:1:1).

Synthesis

Boc-L-Val-(β -OH)-Phe-OH (1). To a precooled solution (-10°C) of Boc-L-Val-OH (3 g, 13.82 mmol) in tetrahydrofuran (THF) (10 ml), *N*-methylmorpholine (NMM) (1.52 ml, 13.82 mmol) and isobutylchloroformate (IBCF) (1.85 ml, 13.82 mmol) were added. After stirring for 10 min, a solution of DL-Phe(β -OH) (2.99 g, 16.56 mmol) in 1 N NaOH (16.5 ml) was added and the mixture stirred at 0°C for 2 h and at room temperature overnight. The organic solvent was removed under reduced pressure and the aqueous phase was acidified with citric acid to pH 3.0 and extracted with ethylacetate (3 \times 15 ml). The ethylacetate layer was washed with water, dried over anhydrous Na₂SO₄ and evaporated to yield (1) as an oily compound. Yield = 4.19 g (80%); $R_F(\alpha) = 0.87$.

Boc-L-Val- Δ Phe-azlactone (2). Compound (1) (2.5 g, 6.57 mmol) was reacted with anhydrous sodium acetate (0.53 g, 6.57 mmol) and freshly distilled acetic anhydride (10 ml) for 24 h at room temperature. The reaction mixture was poured over crushed ice; the resultant precipitate product was washed with 5% NaHCO₃ and water, and finally recrystallized from acetone/water. Yield = 2.3 g (90%); $R_F(\alpha) = 0.93$.



Boc-L-Val- Δ Phe-(β -OH)Phe-OH (3). To a suspension of DL-(β -OH)Phe-OH (1.25 g, 6.97 mmol) in acetone (10 ml) were added with stirring 1 N NaOH (7 ml) and 2 g (5.8 mmol) of azlactone (2). After stirring for 1 h at 45°C , the resulting clear solution was acidified at 0°C by the addition of 1 N HCl (7 ml). The acetone was removed and the aqueous layer was extracted with ethylacetate, washed with water, dried over Na₂SO₄ and evaporated to yield the compound (3).

Boc-L-Val- Δ Phe- Δ Phe-azlactone (4). Peptide (3) (2 g, 3.80 mmol) was reacted with anhydrous sodium acetate (0.31 g, 3.80 mmol) and freshly distilled acetic anhydride (10 ml) for 30 h at room temperature. The reaction mixture was then poured over crushed ice; the resultant precipitate product was washed with 5% NaHCO₃ and water, and finally recrystallized from acetone/water. Yield = 1.8 g (89%).

Boc-L-Val- Δ Phe- Δ Phe-L-Ala-OCH₃ (5). To a solution of (4) (1.0 g, 2.04 mmol) in dichloromethane (DCM) (10 ml), Ala.OMe.HCl (0.41 g, 3.06 mmol) was added, followed by triethylamine (TEA)

Table 1 The Details of Intensity Data Collection and Refinement of *N*-Boc-L-Val- Δ Phe- Δ Phe-L-Ala-OCH₃

Molecular formula	C ₃₂ H ₄₀ N ₄ O ₇
Molecular weight	592.69
Crystal dimensions	0.8 \times 0.3 \times 0.025 mm ³
Crystal system	Hexagonal
Space group	P6 ₁
Z (Molecules/unit cell)	6
<i>a</i> = <i>b</i>	14.912(3) Å
<i>c</i>	25.548(5) Å
<i>V</i>	4912.0(6) Å ³
d.c.	1.217(5) g/cm ³
<i>F</i> (000)	1896
Radiation	CuK α ($\lambda = 1.5418$ Å)
Collected reflections (θ up to 70°)	10,897
R_{sym}	0.054
Unique reflections	3480
Observed reflections ($I > 3\sigma(I)$)	2891
μ_r	0.03
Instrument used	Enraf-Nonius CAD4
Mode of data collection	ω - 2θ
Maximum 2θ	152°
<i>R</i>	0.079
R_w	0.076
<i>S</i>	1.86
Temperature	295 K

Table 2 Atomic Coordinates for the Non-hydrogen Atoms and Their Equivalent Isotropic Thermal Parameters

Atom	X/a	Y/b	Z/c	U_{eq}^a
C ₀₁	1.0088(6)	0.6327(7)	0.0221(4)	0.048(4)
C ₀₂	1.0881(8)	0.6376(12)	0.0174(6)	0.078(6)
C ₀₃	1.0106(9)	0.7344(9)	0.0242(6)	0.070(5)
C ₀₄	1.0266(9)	0.5992(13)	0.0748(5)	0.088(8)
O ₀₁	0.9106(4)	0.5506(4)	0.0001(0)	0.047(2)
C' ₀	0.8188(6)	0.5218(6)	0.0259(4)	0.039(3)
O' ₀	0.8071(5)	0.5670(4)	0.0611(3)	0.051(2)
N ₁	0.7422(5)	0.4372(5)	0.0015(3)	0.045(3)
C ₁ ^α	0.6359(6)	0.3963(6)	0.0170(3)	0.037(3)
C ₁ ^β	0.5651(6)	0.2963(7)	0.0124(3)	0.052(4)
C ₁ ^{γ1}	0.5849(10)	0.2075(8)	0.0050(5)	0.071(5)
C ₁ ^{γ2}	0.4526(8)	0.2659(13)	0.0042(6)	0.089(7)
C' ₁	0.6246(5)	0.3750(5)	0.0776(3)	0.032(3)
O' ₁	0.6577(4)	0.3230(4)	0.0985(3)	0.042(2)
N ₂	0.5756(5)	0.4158(5)	0.1041(3)	0.036(3)
C ₂ ^α	0.5651(5)	0.4064(6)	0.1603(3)	0.033(3)
C ₂ ^β	0.4742(6)	0.3638(6)	0.1858(4)	0.040(4)
C ₂ ^γ	0.3666(6)	0.3081(6)	0.1678(4)	0.041(3)
C ₂ ^{δ1}	0.3324(7)	0.2967(7)	0.1149(4)	0.051(4)
C ₂ ^{δ2}	0.2932(7)	0.2618(7)	0.2077(4)	0.048(4)
C ₂ ^{ε1}	0.2287(7)	0.2375(8)	0.1052(5)	0.059(4)
C ₂ ^{ε2}	0.1883(7)	0.2049(8)	0.1955(5)	0.067(5)
C ₂ ^ζ	0.1546(8)	0.1927(7)	0.1429(5)	0.065(5)
C' ₂	0.6624(6)	0.4473(6)	0.1917(3)	0.041(3)
O' ₂	0.6585(5)	0.4237(5)	0.2375(3)	0.061(3)
N ₃	0.7513(5)	0.5097(5)	0.1668(3)	0.034(3)
C ₃ ^α	0.8497(6)	0.5413(6)	0.1910(3)	0.040(3)
C ₃ ^β	0.9106(7)	0.6351(7)	0.2097(5)	0.057(4)
C ₃ ^γ	0.8964(8)	0.7243(8)	0.2135(5)	0.076(6)
C ₃ ^{δ1}	0.8006(13)	0.7167(11)	0.2138(10)	0.147(12)
C ₃ ^{δ2}	0.9801(13)	0.8180(10)	0.2280(11)	0.156(13)
C ₃ ^{ε1}	0.7910(15)	0.8047(14)	0.2224(8)	0.129(13)
C ₃ ^{ε2}	0.9697(21)	0.9062(14)	0.2284(15)	0.203(20)
C ₃ ^ζ	0.8763(20)	0.8991(15)	0.2191(14)	0.208(19)
C' ₃	0.8893(6)	0.4685(7)	0.1906(4)	0.044(3)
O' ₃	0.9705(4)	0.4867(7)	0.2123(3)	0.050(3)
N ₄	0.8332(6)	0.3797(5)	0.1618(3)	0.048(3)
C ₄ ^α	0.8598(8)	0.2987(7)	0.1597(4)	0.057(5)
C ₄ ^β	0.7647(10)	0.1938(9)	0.1699(6)	0.076(6)
O' ₄	0.9655(8)	0.2631(9)	0.1047(5)	0.118(7)
C' ₄	0.9051(7)	0.2963(8)	0.1064(5)	0.061(5)
O ₂	0.8673(7)	0.3206(7)	0.0682(4)	0.082(5)
C ₅	0.8966(14)	0.3080(16)	0.0148(5)	0.095(10)

$$^a U_{eq} = 1/3 \sum_i \sum_j U_{ij} a_i^* a_j^* a_i a_j.$$

(0.33 ml, 2.45 mmol) and the mixture was stirred at room temperature for 40 h. The solvent was removed *in vacuo*, the organic residue was dissolved in ethylacetate and washed with 10% NaHCO₃, 5% citric acid and water and dried over anhydrous

Na₂SO₄. The solvent was removed, the solid product was collected and recrystallized from acetone/water and THF/petroleum ether. Yield = 0.8 g (63.4%); $R_F(a) = 0.72$, $R_F(b) = 0.923$; melting point, m.p., = 197°C.

Structure Determination

Single crystals of peptides were obtained by slow evaporation of its solution in an acetone/water mixture (70:30) at room temperature. The intensity data were collected on a Nonius CAD4 diffractometer equipped with a graphite monochromator and with CuK α radiation in the ω -2 θ (θ up to 76 $^\circ$) scanning mode. The absorption was disregarded ($\mu_r=0.03$) and data were corrected for Lorentz and polarization factors. The crystallographic data are listed in Table 1. The structure was determined by direct methods using SHELXS86 [7]. All the non-hydrogen atoms were refined anisotropically using a full-matrix structure factor least-squares procedure using $|F|$ values (SHELX76) [8]. The positions of hydrogen atoms were determined by difference Fourier calculations. The hydrogen atoms were refined using isotropic thermal parameters. The atomic scattering factors used in these calculations were those of Cromer and Mann [9] for non-hydrogen atoms and of Stewart *et al.* [10] for hydrogen atoms. The final R factor for 2891 observed ($I \geq 3\sigma$) reflection is 0.079. The final positional and equivalent isotropic thermal parameters of non-hydrogen atoms are given in Table 2.

RESULTS AND DISCUSSION

Molecular Dimensions

The introduction of a double bond between the C $^\alpha$ and C $^\beta$ atoms in Δ Phe residues affects the other bond lengths and angles in the same residues as seen in other Δ Phe-containing peptides. These distances in Δ Phe 2 and Δ Phe 3 residues are 1.34(1) Å and 1.32(1) Å respectively and correspond to a classical C=C double bond distance of 1.337 Å [11]. The N $_2$ - C $_2^\alpha$ =1.44(2) Å, C $_2^\alpha$ - C $_2'$ =1.49(1) Å, N $_3$ - C $_3^\alpha$ =1.44(1) Å, C $_3^\alpha$ - C $_3'$ =1.47(2) Å bond distances in Δ Phe 2 and Δ Phe 3 residues are slightly shorter than the corresponding bond distances in saturated residues (1.45 Å for N - C $^\alpha$ and 1.53 Å for C $^\alpha$ - C') [12]. The shortening of the bonds N $_2$ - C $_2^\alpha$, C $_2^\alpha$ - C $_2'$, N $_3$ - C $_3^\alpha$ and C $_3^\alpha$ - C $_3'$ is probably due to the sp 2 hybridized C $_2^\alpha$, C $_2^\beta$, C $_3^\alpha$ and C $_3^\beta$ atoms and might also be a result of partial conjugation of Δ Phe ring electrons and remaining atoms in the residue. As indicated by the torsion angles ($\chi_2^1=7(1)$, $\chi_2^{2,1}=-8(2)$, $\chi_2^{2,2}=170(1)$, $\chi_3^1=3(2)$, $\chi_3^{2,1}=23(2)$, $\chi_3^{2,2}=-170(1)^\circ$), the Δ Phe rings and peptide units are found to be coplanar in the structure. The values of bond angles N $_2$ - C $_2^\alpha$ - C $_2'$ and N $_3$ - C $_3^\alpha$ - C $_3'$ are

117(1) $^\circ$ and 118(1) $^\circ$ respectively, which are slightly less than the standard trigonal values of 120 $^\circ$, while the bond angles N - C $_2^\alpha$ - C $_2^\beta$, N - C $_3^\alpha$ - C $_3^\beta$, C $_2^\alpha$ - C $_2^\beta$ - C $_2^\gamma$ and C $_3^\alpha$ - C $_3^\beta$ - C $_3^\gamma$ with values of 124(1) $^\circ$, 123(1) $^\circ$, 132(1) $^\circ$ and 131(1) $^\circ$ respectively are considerably larger than 120 $^\circ$. Because of the shortening of the distance between the C $^\alpha$ and C $^\beta$ atoms and the enhanced planarity in the dehydro-residue, the side-chain atoms approach the backbone atoms. This leads to some unfavourable interactions, thus causing a rearrangement of the bond angles at the C $^\alpha$ and C $^\beta$ atoms which are manifested in the above-mentioned derivations from the standard values. The remaining bond lengths and angles are normal.

Conformation of the Peptide

The perspective stereoview of the molecule with numbering scheme is shown in Figure 1. The selected torsion angles are listed in Table 3. The backbone torsion angles $\phi_1=-54(1)^\circ$, $\psi_1=129(1)^\circ$, $\omega_1=-177(1)^\circ$, $\phi_2=57(1)^\circ$, $\psi_2=15(1)^\circ$, $\omega_2=-170(1)^\circ$, $\phi_3=80(1)^\circ$, $\psi_3=7(2)^\circ$, $\omega_3=-177(1)^\circ$, $\phi_4=-108(1)^\circ$ and $\psi_4^T=-34(1)^\circ$ indicate that the peptide adopts a folded structure with two overlapping respectively. These turns are stabilized by two intramolecular hydrogen bonds between the CO of the Boc group and the NH of Δ Phe 3 , and the CO of Val 1 and the NH of Ala 4 . Thus the Δ Phe 2 is located at the ($i+2$) position of a β -turn II and at the ($i+1$) position of a β -turn III'. The second dehydro-residue Δ Phe 3 is located at the ($i+2$) position of a β -turn III'. As seen from Table 3, the torsion angles of the valine side chain correspond to the most frequently observed values for a valyl residue [13]. The torsion angles of Δ Phe 2 and Δ Phe 3 side chains are similar and indicate that the two Δ Phe residues are essentially planar.

As seen from Table 4, it is noteworthy that the conformation of the present peptide has been found to be similar to that observed for Boc-Val- Δ Phe- Δ Phe-Val-OCH $_3$ [6] with two overlapping β -turns of types II and III' respectively, whereas it is slightly different from the one found in Boc-Ala- Δ Phe- Δ Phe-NHCH $_3$ [5] which contains two overlapping β -turns of type III. These observations suggest that two consecutive Δ Phe residues substituted at ($i+2$) and ($i+3$) positions with branched β -carbons such as in Val and Ile at both ends, as well as a single Val/Ile placed on one side, generates folded structures with two overlapping β -turns of types II and III' respectively. On the other hand, the substitution of non-C $^\beta$ -branched residues on both sides of a Δ Phe- Δ Phe

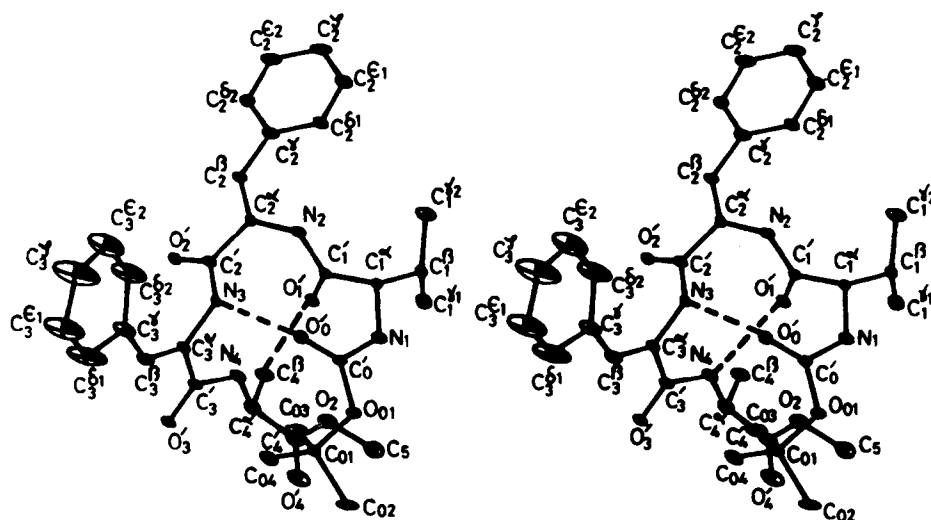


Figure 1 Perspective stereoview of the molecule and the numbering scheme. The dashed lines indicate hydrogen bonds.

Table 3 Selected Torsion Angles

θ_0	$C_1 - O_1 - C'_0 - N_1$	-173(1)
ω_0	$O_1 - C'_0 - N_1 - C_1^\alpha$	-174(1)
ϕ_1	$C'_0 - N_1 - C_1^\alpha - C'_1$	-54(1)
ψ_1	$N_1 - C_1^\alpha - C'_1 - N_2$	129(1)
χ_1	$N_1 - C_1^\alpha - C_1^\beta - C_1^{\gamma 1}$	60(1)
$\chi_1^{2.1}$	$N_1 - C_1^\alpha - C_1^\beta - C_1^{\gamma 2}$	-168(1)
ω_1	$C_1^\alpha - C'_1 - N_2 - C_2^\alpha$	-177(1)
ϕ_2	$C'_1 - N_2 - C_2^\alpha - C'_2$	57(1)
ψ_2	$N_2 - C_2^\alpha - C'_2 - N_3$	15(1)
χ_2^1	$N_2 - C_2^\alpha - C_2^\beta - C_2^\gamma$	7(1)
$\chi_2^{2.1}$	$C_2^\alpha - C_2^\beta - C_2^\gamma - C_2^{\delta 1}$	-8(2)
$\chi_2^{2.2}$	$C_2^\alpha - C_2^\beta - C_2^\gamma - C_2^{\delta 2}$	170(1)
ω_2	$C_2^\alpha - C'_2 - N_3 - C_3^\alpha$	-171(1)
ϕ_3	$C'_2 - N_3 - C_3^\alpha - C'_3$	80(1)
ψ_3	$N_3 - C_3^\alpha - C'_3 - N_4$	7(2)
χ_3^1	$N_3 - C_3^\alpha - C_3^\beta - C_3^\gamma$	3(2)
$\chi_3^{2.1}$	$C_3^\alpha - C_3^\beta - C_3^\gamma - C_3^{\delta 1}$	23(2)
$\chi_3^{2.2}$	$C_3^\alpha - C_3^\beta - C_3^\gamma - C_3^{\delta 1}$	-170(1)
ω_3	$C_3^\alpha - C'_3 - N_4 - C_4^\alpha$	-177(1)
ϕ_4	$C'_3 - N_4 - C_4^\alpha - C'_4$	-108(1)
ψ_4^T	$N_4 - C_4^\alpha - C'_4 - O_2$	-34(1)

sequence results in the formation of a folded structure with two overlapping β -turns of type III (incipient 3_{10} -helix). Thus, the present dehydropolypeptide structure has established that a 3_{10} -helix can only be formed in a tetrapeptide if the $-\Delta$ Phe- Δ Phe-sequence does not have branched β -carbon residues on either side of it. However, if a folded structure with two overlapping β -turns of types II and III' is to be generated, the sequence must have Val or Ile on both sides or at least on one side of the Δ - Δ Phe segment in a tetrapeptide. These observations can be exploited for a useful design.

Molecular Packing and Hydrogen Bonding

The molecular packing in the unit cell as viewed along the a -axis is shown in Figure 2. The axis of the folded structure is approximately parallel to the crystallographic c -axis. The NH and C=O groups which are not involved in intramolecular hydrogen bonds are positioned at the opposite ends of the folded molecules. Therefore, the molecules form

Table 4 The ϕ, ψ Torsion Angles in (I) Boc-Ala- Δ Phe- Δ Phe-NHCH₃^a, (II) Boc-Val- Δ Phe- Δ Phe-Val-OCH₃, (III) Boc-Val- Δ Phe- Δ Phe-Ala-OCH₃

Peptides	ϕ_1	ψ_1	ϕ_2	ψ_2	ϕ_3	ψ_3	ϕ_4	ψ_4^T
I	-71.0(4)	-25.0(4)	-63.1(4)	-11.5(4)	-62.4(3)	-24.2(4)	-	-
	37.1(5)	59.7(4)	67.6(4)	6.6(4)	59.9(4)	25.1(5)	-	-
II	-56.5(4)	130.5(4)	65.8(5)	12.8(6)	79.4(6)	3.9(7)	-106.4(5)	-54.6(6)
III	-54(1)	129(1)	57(1)	15(1)	80(1)	7(2)	-108(5)	-34(1)

^aTwo molecules in the asymmetric unit.

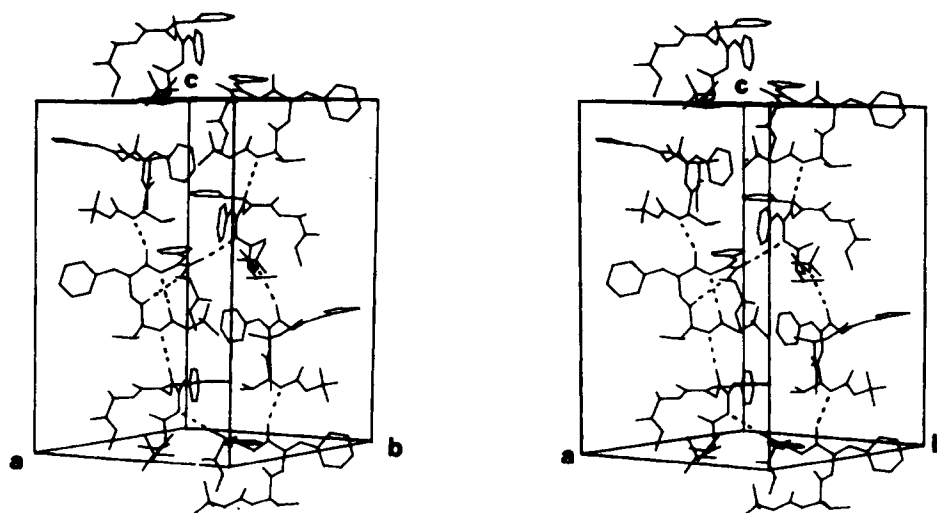


Figure 2 Stereoview of the molecular packing in unit cell. The dashed lines indicate hydrogen bonds.

Table 5 The Parameters of Hydrogen Bonds

D-H...A	D-H (Å)	H...A (Å)	D...A (Å)	Angle (°)	Symmetry
N ₃ -H ₃ ...O' ₀	0.8(1)	2.1(1)	2.83(1)	165(1)	<i>x,y,z</i>
N ₄ -H ₄ ...O' ₁	1.1(1)	1.8(1)	2.83(1)	155(1)	<i>x,y,z</i>
N ₁ -H ₁ ...O' ₂	0.7(1)	2.3(1)	2.97(1)	144(1)	$-x+y+1, -x+1, z-1/3$
N ₂ -H ₂ ...O' ₃	1.1(1)	1.8(1)	2.87(1)	161(1)	$y, -x+y+1, z-1/6$

head-to-tail hydrogen bonds giving rise to continuous helical columns which run parallel to the *c*-axis. The helical columns are arranged in a hexagonal packing mode. The details of hydrogen bond parameters are given in Table 5.

CONCLUSIONS

The conformations of peptides containing two consecutive Δ Phe residues indicate the following:

1. A three peptide unit sequence with two consecutive Δ Phe residues substituted at (*i*+1) and (*i*+2) positions results in the formation of an S-shaped structure with positive and negative ϕ, ψ torsion angles alternately.
2. A tetrapeptide with two consecutive Δ Phe residues at (*i*+2) and (*i*+3) positions adopts folded structures.
3. A tetrapeptide containing two consecutive Δ Phe residues at (*i*+2) and (*i*+3) positions with residues other than the branched β -carbons at (*i*+1) and (*i*+4) positions forms two overlapping type III β -turns (incipient 3₁₀-helix).
4. A sequence containing two consecutive Δ Phe residues at (*i*+2) and (*i*+3) positions with branched β -carbon residues at (*i*+1) and (*i*+4) positions adopts a conformation with two overlapping β -turns of types II and III' respectively.
5. A sequence with a branched β -carbon residue only on the one side of a $-\Delta$ Phe- Δ Phe- segment also assumes a conformation with two overlapping β -turns of types II and III'.

Acknowledgements

The authors thank the Council of Scientific and Industrial Research, New Delhi, for financial support. Crystallographic data (excluding structure factors) for the structure reported in this paper have been deposited with the Cambridge Crystallographic Data Center as a supplementary publication. Copies of the data can be obtained, free of charge, on application to the Director, CCDC, 12 Union Road, Cambridge CB2 1EZ, UK. Fax +4 1223 336 033 or email: teched@chemcrys.cam.ac.uk.

REFERENCES

1. T. P. Singh, P. Narula and H. C. Patel (1990). α,β -Dehydro-residues in the design of peptide and protein structures. *Acta Crystallogr. B* 46, 539–000.
2. T. P. Singh, B. Padmanabhan, P. Narula, A. K. Saxena, Ch. Betzel, P. Sharma and S. Dey in: *Proceedings of the International Symposium on Practical Protein Engineering on Subtilisin Enzymes*, R. Bott and Ch. Betzel, Eds., p. 20–24, Plenum Press, New York 1994.
3. O. Pieroni, A. Montagnoli, A. Fissi, S. Merlino and F. Ciardelli (1975). Structure and optical activity of unsaturated peptides. *J. Am. Chem. Soc.* 97, 6820–6826.
4. O. Pieroni, A. Fissi, S. Merlino and F. Ciardelli (1976/1977). Chiroptical properties and conformation of Δ -phenylalanine peptides. *Israel J. Chem* 15, 22–28.
5. A. Tuzi, M. R. Ciajolo, G. Guarino, P. A. Temussi, A. Fissi and O. Pieroni (1993). Solid state and solution structure of Boc-L-Ala- Δ Phe- Δ Phe-NHMe: a dehydro-peptide showing propensity for 3_{10} -helices of both screw senses. *Biopolymers* 33, 1111–1121.
6. S. Dey, S. N. Mitra and T. P. Singh (1995). Design of peptides using α,β -dehydro-residues: synthesis, crystal structure and molecular conformation of N-Boc-Val- Δ Phe- Δ Phe-Val-OCH₃. *Biopolymers* In press.
7. G. M. Sheldrick: *SHELXS86. A Program for Crystal Structure Determination*. Anorganisch-Chemisches, Institut der Univ. Gottingen, Germany 1986.
8. G. M. Sheldrick: *SHELX76. A Program for Crystal Structure Determination*. Anorganisch-Chemisches, Institut der Univ. Gottingen, Germany 1976.
9. D. T. Cromer and J. B. Mann (1968). X-ray scattering factors computed from numerical Hartree-Fock wave functions. *Acta Crystallogr. A* 24, 321–324.
10. R. F. Stewart, E. R. Davidson and W. T. Simpson (1965). Coherent X-ray scattering for the hydrogen atom in the hydrogen molecule. *J. Chem. Phys.* 42, 3175–3187.
11. R. E. Dickerson and I. Geis in: *The Structure and Action of Proteins*, p. 13, Harper & Row, New York 1969.
12. E. Benedetti in: *Peptides, Proceedings of the Fifth American Peptide Symposium*, M. Goodman and J. Meinhofer, Eds., p. 257–273, Wiley, New York 1977.
13. E. Benedetti, G. Morelli, G. Nemethy and H. A. Scheraga (1983). Statistical and energetic analysis of side-chain conformations in oligopeptides. *Int. J. Peptide Protein Res.* 22, 1–15.